# Bacterial DNA extraction from raw milk using Ribospin™ Pathogen/TNA kit

## **Experimental Conditions**

#### **Materials Required**

- Ribospin™ Pathogen/TNA (50 preps: 341-150 / 250 preps: 341-152)
- Absolute ethanol (C<sub>2</sub>H<sub>6</sub>O, CAS No.: 64-17-5, ≥99.0%)
- 1.5 ml or 2.0 ml microcentrifuge tube
- Vortex mixer
- Centrifuge (Max. speed 14,000 rpm or ≥10,000 x g)
- · Pipette & sterile pipette tips
- Suitable protector (e.g., lab coat, disposable gloves, goggles, etc.)

## **Sample Information**

 Sample type: raw milk (with the spiked vaccine of Mycoplasma spp.)



- · Sampling: after collecting fresh raw milk samples, immediately inject the vaccines of Mycoplasma spp. (10° to 10-4 dilution) in the samples and extract the bacterial DNA.
- Extraction conditions
  - Sample amount: 1 ml
  - Elution volume: 50 μl

## **Protocol**

#### **Before Starting**

- Before using for the first time, add absolute ethanol (ACS grade or better) into Buffer RB1, Buffer RBW, and Buffer RNW as indicated on the bottle.
- If a precipitate has formed in Buffer KL, heat to dissolve at 56 °C before use.

### **Sample Preparation**

- 1. Transfer 1.0 ml of the raw milk to a 1.5 or 2.0 ml microcentrifuge
- 2. Centrifuge at 10,000 x g above for 5 min at room temperature and discard the supernatant containing fat and liquid layer.
- 3. The next step is according to **Ribospin™ Pathogen/TNA protocol**.

## Ribospin™ Pathogen/TNA Protocol (Raw Milk)

\* For more details and methods, please refer to the handbook of Ribospin™ Pathogen/TNA.

Lysis	<ol> <li>Add 200 μl Buffer SL and resuspend the pellets.</li> <li>Add 20 μl of Proteinase K solution and 200 μl of Buffer KL. Vortex vigorously and incubate at 20-25 °C for 5 min.</li> </ol>
Binding	<ol> <li>Add 300 μl Buffer RB1, pulse-vortex to mix, and spin down briefly.</li> <li>Transfer the mixture to Column Type P and centrifuge at 10,000 x g above for 1 min.</li> </ol>
Washing	<ol> <li>Add 600 µl Buffer RBW to Column Type P and centrifuge at 10,000 x g above for 1 min.</li> <li>Add 600 µl Buffer RNW to Column Type P and centrifuge at 10,000 x g above for 1 min.</li> <li>Centrifuge at full speed for 1 min to remove the residual wash buffer.</li> <li>Transfer Column Type P to a new 1.5 ml microcentrifuge tube.</li> </ol>
Elution	<ol> <li>Add 50 μl of Nuclease-free Water to Column Type P and incubate at room temperature for 1 min.</li> <li>Centrifuge at full speed for 1 min.</li> </ol>

Table 1. Brief protocol of GeneAll® Ribospin™ Pathogen/TNA kit for bacterial DNA purification

#### Result

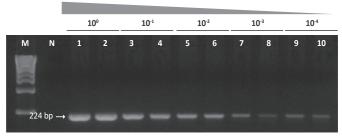


Figure 1. Conventional PCR amplification result of Mycoplasma spp. gene using DNA extracted from raw milk as template

M: GENESTA™ 1 kb DNA ladder with 5X loading dye (GA-100) N: negative control (nuclease-free water)

Lanes 1, 2: amplified bacterial DNA

Lanes 3, 4: amplified bacterial DNA (10<sup>-1</sup> dilution)

Lanes 5, 6: amplified bacterial DNA (10<sup>-2</sup> dilution) Lanes 7, 8: amplified bacterial DNA (10<sup>-3</sup> dilution)

Lanes 9, 10: amplified bacterial DNA (10-4 dilution)

※ Electrophoresis conditions: 2% agarose gel, 150 V, 15 min, 2 μl loading